



Preliminary phytochemical screening and antitrichomonal activity of Ferula pseudalliacea

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Abstract

Background: *Trichomonas vaginalis* (*T. vaginalis*) causes human trichomoniasis, a common type of protozoan vaginitis. Due to the increasing incidence of drug-resistant trichomoniasis, new pharmacological research is needed. The aim was to investigate the activity of *Ferula pseudalliacea* (*F. pseudalliacea*) against *T. vaginalis* and to perform a preliminary phytochemical analysis of its extracts.

Methods: Essential oil and various extracts of *F. pseudalliacea* roots, including n-hexane, ethyl acetate, and methanol, were obtained. Susceptibility testing of the plant products was performed on five *T. vaginalis* isolates using the microtiter plate method. Minimum lethal concentration (MLC) and growth inhibitory percent (GI%) of sub-MLC concentrations were reported after 24- and 48-hour exposures. Phytochemical screening of the extracts was carried out using a standard procedure.

Results: The antitrichomonal effect of the plant products depended on time and concentration, with the greatest effect observed after 48 hours of exposure. The essential oil and n-hexane extract of F pseudalliacea demonstrated remarkable activity with MLC of 250 μ g/ml, followed by the ethyl acetate (MLC=500 μ g/ml) and methanol extract (MLC=1000 μ g/ml), with GI% 92.8, 50.6, 85.2, and 42.8, respectively. The bioactive constituents of the extracts were coumarins, terpenoids, steroids, phenols, tannins, and glycosides.

Conclusion: The results of this study demonstrated in vitro antitrichomonal properties of *F. pseudalliacea*. Therefore, further studies are needed to investigate the potential antitrichomonal activity of its bioactive constituents.

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Introduction

Trichomoniasis is a common cause of human vaginitis caused by the protozoan parasite *Trichomonas vaginalis*. The infection is one of the most prevalent STIs worldwide. According to the World Health Organization in 2016, there were 156 million new cases of *Trichomonas* infection among persons aged 15 to 49 years. Trichomoniasis in women may present with a wide variety of clinical features, from an asymptomatic infection to severe vaginitis. Adverse pregnancy outcomes, infertility, and cervical neoplasia may be seen as complications after trichomoniasis. Furthermore, the infection increases the risk of spreading HIV infection in the community (1,2).

Metronidazole therapy is commonly the standard treatment for trichomoniasis worldwide. Since 1961, when metronidazole was introduced to treat trichomoniasis, it has faced challenges. One of these is drug resistance. Drug-resistant *T. vaginalis* is involved in an increasing number of refractory trichomoniasis cases. The first metronidazole-resistant trichomoniasis was reported in 1962, and it has been on the rise. According to the Center for Disease Control and Prevention, 2 to 5% of clinical *T. vaginalis* isolates are metronidazole-resistant in the United States (3,4).

Due to the presence of bioactive compounds in plants, they are of particular interest in pharmaceutical research. Plant essential oils and extracts are known to be a rich source of natural ingredients for the treatment of various diseases and are more compatible with biological systems. Ferula is a genus of plants with about 185 species. This genus has a special position in the Apiaceae family because of its pharmaceutical and industrial importance. In the Iranian flora, the genus Ferula contains 32 species, including 15 endemic plants, and this genus is typically called koma or kema. Several species of Ferula have been used in folk remedies as treatment of stomachache, hysteria, arthritis,

rheumatoid disorders, and etc. Recent studies have proven antibacterial, antileishmanial, antimalarial, antioxidant, anti-epileptic, and anti-inflammatory effects of *Ferula* species (5-7). In previous studies, the methanol extract of *F. szowitsiana* was able to inhibit the growth of *T. vaginalis* cells (8).

F. pseudalliacea is an indigenous species of Iran and grows in the Sanandaj Mountains (West of Iran), and their gum has been used in traditional medicine for healing wounds and relieving itching. Recently, different studies have been done on the antibacterial, antiplasmodial, phytotoxic, and anticancer activity of F. pseudalliacea (9-12). But so far, a study to evaluate the antitrichomonas effect of this species has not been done.

In this study, we have examined the antitrichomonas effect of *F. pseudalliacea* essential oil and extracts against *T. vaginalis*. Also, the preliminary phytochemical analysis of the extracts was performed by standard methods.

Methods

Plant material

The roots of *F. pseudalliacea* were obtained from natural habitats in western Iran. The plant was identified in the herbarium of the Department of Pharmacognosy of Hamadan University of Medical Sciences, and voucher number 234 was allocated.

Preparation of the essential oil, extract, and phytochemical screening

The plant was dried using the shade-drying method at room temperature $(20\pm5~^\circ\text{C})$, and the dried plant materials were crushed into powder. The powder (100~g) was used to prepare the essential oil using a Clevenger-type apparatus. The obtained essential oil was kept in an airtight container in a refrigerator $(4~^\circ\text{C})$ until use. Extraction of the dried plant

was performed using the maceration method. Briefly, the powdered plant (100 g) was macerated separately in n-hexane, ethyl acetate, and methanol solvents (3 \times 2 L, room temperature for 72 h, 25 °C). Extraction was performed using a rotary evaporator below 40 °C. The obtained extracts were kept in dark containers in a refrigerator (4 °C) until use. Phytochemical analysis of the extracts was done using standard methods, and their constituent compounds were identified according to the method of Ugochukwu and Bargah (13,14).

Parasite culture and solutions

Five clinical *T. vaginalis* isolates were cultured in TYI-S-33 medium supplemented with 10% heat-inactivated adult bovine serum and antibiotics (100 IU/ml penicillin and 100 μ g/ml streptomycin). After several 48-hour subcultures, pure trophozoites in the log phase of growth were used for the susceptibility assay (15,16). Metronidazole (Sigma-Aldrich, St Louis, USA) was dissolved in distilled water. The plant products were dissolved in dimethyl sulfoxide (D2650 SIGMA, BioReagent) or distilled water according to solubility. Solubility of the essential oil and the extracts in the culture medium was considered as the criterion for determining plant product concentrations, and susceptibility testing was started with the highest concentration. Solutions were prepared in 2-fold dilutions in culture medium for susceptibility assays at the following concentrations: 200, 100, 50, 25, 12.5, 6.2, 3.1, 1.6, 0.8, 0.4, 0.2, 0.1 μ g/ml for metronidazole, and 4000, 2000, 1000, 500, 250, 125, 62.5 μ g/ml for the plant products.

Susceptibility assay

The minimum lethal concentration (MLC) corresponds to the lowest concentration of the antitrichomonal agents that kills all trophozoites after exposure. Growth inhibition due to sub-MLC and lower concentrations of the agents was considered as the percentage of growth inhibition (GI% = a - b / a \times 100; a = mean number of viable trophozoites in the negative control well, b = number of viable trophozoites in the test well at \leq sub-MLC concentration). Susceptibility testing was assessed according to the method recommended by the CDC (17).

The experiments were performed in 96-well microtiter plates. First, the required serial dilutions of the agents were prepared, and 100 μL of the prepared solutions was dispensed into the wells. Then, the parasites in the logarithmic growth phase were counted with a hemocytometer (Neubauer cell chamber), and 100 μL of parasite-containing medium (2 \times 10s trophozoites/mL) was added to each test well. Finally, the number of *Trichomonas* cells was set to 2 \times 10s cells/well. The plates were aerobically incubated at 35.5 °C. After 24 and 48 hours of exposure, the

test plates were examined with an inverted microscope to determine the MLC. The lowest concentration of the essential oil and extracts in the test well where no motile parasites were observed was considered the MLC concentration.

To evaluate the GI% rate of the agents, the number of parasites in the test wells was counted and compared with the number of parasites in the negative control well according to the equation mentioned above. The experiments were repeated in pairs and twice separately under sterile conditions. In each run, control wells (Negative control and metronidazole control) were used to check the experimental conditions. At the end, exposed parasites were cultured in fresh medium for MLC confirmation.

Statistical analysis

Analysis was performed using SPSS statistical software, version 16. The results were shown as MLC and mean values. The Friedman test was used to compare the averages. A P value less than 0.05 was considered statistically significant.

Results

Susceptibility testing revealed that the extracts and essential oil of F. pseudalliacea had a lethal effect on the Trichomonas parasite. The antitrichomonal activity of F. pseudalliacea depended on concentration and time of exposure, as shown in Tables 1 and 2. At MLC concentrations, the tested agents were able to kill all trophozoites, which was confirmed by culturing treated trophozoites in fresh medium. The essential oil of F. pseudalliacea exhibited the highest anti-Trichomonas potential with an MLC of 250 µg/ml, followed by the ethyl acetate and n-hexane extracts with MLC of 500 μg/ml, after 24 hours of incubation (P=0.002). After 48 hours, the antitrichomonal activity of the n-hexane extract increased to an MLC of 250 µg/ml (Table 2). At the sub-MLC concentrations, growth inhibition of the trichomonads was observed, and the number of live trophozoites was remarkably reduced compared to the control (Tables 1 and 2). GI% of the agents at the sub-MLC concentrations ranged from 35.2% to 87.0% after 24 hours, and from 42.8% to 92.8% after 48 hours. Drug susceptibility testing demonstrated that the Trichomonas isolates were susceptible to metronidazole, with MLCs ranging from 6.2 to 12.5 µg/ml (Table 3).

Preliminary phytochemical tests for extracts were studied by standard methods. The results showed *F. pseudalliacea* extracts contain bioactive constituents including coumarins, terpenoids, steroids, phenolics, tannins, and glycosides (Table 4). The major constituents were coumarins, terpenoids, and steroids.

Table 1. Efficacy of essential oil and extracts of F. pseudalliacea on T. vaginalis after 24 hours' exposure

Agents	Mean and standard deviation of growth inhibition percent (GI%) at different concentrations of plant products								
	62.5 (μg/ml)	125 (μg/ml)	250 (µg/ml)	500 (μg/ml)	1000 (μg/ml)	2000 (μg/ml)	4000 (μg/ml)		
Essential oil	47.6±5.6	59.6±7.3 b	100.0±0.0 a	100.0±0.0	100.0±0.0	100.0±0.0	100.0±0.0		
N-hexane extract	0.0 ± 0.0	35.0±4.5	74.2±2.6 b	100.0±0.0 a	100.0±0.0	100.0±0.0	100.0±0.0		
Ethyl acetate extract	0.0 ± 0.0	0.0 ± 0.0	35.2±2.4 b	100.0 a ±0.0 a	100.0±0.0	100.0±0.0	100.0±0.0		
Methanol extract	0.0 ± 0.0	0.0 ± 0.0	0.0 ± 0.0	34.2±3.7	87.0±1.6 b	100.0±0.0 a	100.0±0.0		

a Minimum Lethal Concentration (MLC) is related to the lowest concentration of the antitrichomonal agents that kill all trichomonads

Table 2. Efficacy of essential oil and extracts of F. pseudalliacea on T. vaginalis after 48 hours' exposure

Agents	Mean and standard deviation of growth inhibition percent (GI%) at different concentrations of plant products									
	Agents	62.5 (µg/ml)	125 (μg/ml)	250 (μg/ml)	500 (μg/ml)	1000 (μg/ml)	2000 (μg/ml)	4000 (μg/ml)		
	Essential oil	62.4±7.7	92.8±1.9 b	100.0±0.0 a	100.0±0.0	100.0±0.0	100.0±0.0	100.0±0.0		
]	N-hexane extract	0.0 ± 0.0	50.6±2.1 b	100.0°±0.0°	100.0±0.0	100.0±0.0	100.0±0.0	100.0±0.0		
Et	thyl acetate extract	0.0 ± 0.0	0.0 ± 0.0	85.2±2.2 b	100.0±0.0 a	100.0±0.0	100.0±0.0	100.0±0.0		
	Methanol extract	0.0 ± 0.0	0.0 ± 0.0	0.0 ± 0.0	42.8±1.9 b	100.0±0.0 a	100.0±0.0	100.0±0.0		

^a Minimum Lethal Concentration (MLC) is related to the lowest concentration of the antitrichomonal agents that kill all trichomonads

Table 3. Efficacy of metronidazole on *T. vaginalis*

Incubation time	Mean and standard deviation of growth inhibition percent (GI%) at different concentrations of metronidazole										
incubation time	0.4 (µg/ml)	0.8 (µg/ml)	1.6 (µg/ml)	3.1 (µg/ml)	6.2 (µg/ml)	12.5 (μg/ml)	25 (μg/ml)	50 (μg/ml)	100 (μg/ml)	200 (μg/ml)	
24 hours	51.8±2.4	59.0±5.6	71.8±2.6	83.6±2.5	95.4±0.5 b	100.0±0.0 a	100.0±0.0	100.0±0.0	100.0±0.0	100.0±0.0	
48 hours	63.0±2.9	82.4±2.1	93.5±1.3	98.6±0.5 b	100.0±0.0 a	100.0±0.0	100.0±0.0	100.0±0.0	100.0±0.0	100.0±0.0	

^a Minimum Lethal Concentration (MLC) is related to the lowest concentration of the antitrichomonal agents that kill all trichomonads

^b Sub-MLC concentration is related to inhibition of trichomonads growth

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^b Sub-MLC concentration is related to inhibition of trichomonads growth

Table 4. Preliminary phytochemical screening of F. pseudalliacea extracts

Compounds	Test methods	Result				
Compounds	Test methods	n-hexane	ethyl acetate	methanol		
Carbohydrates	Fehling's solutions	-	-	-		
Glycosides	Keller-kilani	-	-	+		
Pheolics	Ferric chloride	+	+	2+		
Tannins	Ferric chloride	-	-	+		
Alkaloids	Dragendorff's	-	-	-		
Proteins and amino acids	Ninhydrin test	-	-	-		
Coumarins	UV test	3+	3+	3+		
Saponins	Foam test	-	-	-		
Flavonoids	Alkaline reagent	-	-	-		
Phlobatannins	Precipitate test	-	-	+		
Terpenoids	-	3+	3+	2+		
Steroids	Salkowski,s test	2+	2+	2+		

+ Presence: - Absence

Discussion

In this research, the antitrichomonal efficacy of F, pseudalliacea was evaluated in comparison with metronidazole, the first-line treatment for trichomoniasis. The results showed that F, pseudalliacea was potentially effective against T, vaginalis. The essential oil and n-hexane extract of F, pseudalliacea were the most potent antitrichomonal agents. After 48 hours of exposure, the MLC of these two potent products was 250 μ g/ml. At sub-MLC concentrations, the GI% of the oil and the n-hexane extract was 92.8% and 50.6%, respectively.

To our knowledge, the antimicrobial activity of essential oil and crude extracts of *F. pseudalliacea* has not been previously investigated. However, the in vitro antiplasmodial and antibacterial activities of coumarin derivatives from *F. pseudalliacea* have been demonstrated. Anti-*Plasmodium falciparum* activity of sanandajin, methyl galbanate, and kamolonol acetate was reported with IC50 values of 2.6, 7.1, and 16.1 μM, respectively (9). Sanandajin and ethyl galbanate were effective against *Staphylococcus aureus* and *Helicobacter pylori* (MIC=64 μg/ml), and methyl galbanate was effective against a vancomycinresistant strain of *Enterococcus faecium* (MIC=64 μg/ml) (12).

To date, many studies have explored the antimicrobial effects of medicinal plants, some specifically targeting the *Trichomonas* parasite. Twenty-six Iranian medicinal plants with antitrichomonal activity were reviewed by Ziaei Hezarjaribi et al. (18). In this review, *Artemisia aucheri*, *Zataria multiflora*, and *Lavandula angustifolia* were highlighted as the most potent medicinal plants. Other Iranian medicinal plants with strong antitrichomonal activity include *Foeniculum vulgare*, *Marrubium vulgare*, *Pistacia atlantica* subsp. *kurdica*, *Plantago lanceolata* L., and *Ferula* gummosa.

The antitrichomonal activity of *F. vulgare* was investigated by Karami et al. In their study, the methanolic and hexanic extracts of the medicinal herb showed the greatest potency with an MLC of 360 μg/ml. The essential oil demonstrated lower activity (MLC=1600 μg/ml). Chemical analysis of the essential oil revealed that E-anethole was the major component (19). In another study by Akbari et al., the essential oil of *M. vulgare* displayed the highest activity against *T. vaginalis* (MLC=291 μg/ml), while the n-hexane extract showed the lowest activity (MLC=1500 μg/ml) (20).

Matini and colleagues evaluated the efficacy of *P. atlantica* subsp. *kurdica* and *P. lanceolata L.* extracts against *T. vaginalis*. The ethyl acetate extract of these plants was considered the most potent antitrichomonal product, with MLC values of 337 and 1525 μ g/ml, respectively (21,22). Anti-*Trichomonas* properties of various extracts of *F. gummosa*-including ethyl acetate, n-hexane, methanol, and its essential oil-were examined by Akbari et al. Their results showed that the extracts were more effective (MLC=125 μ g/ml) than the essential oil (MLC=500 μ g/ml). α - and β -Pinene, and β -eudesmol were identified as the major components of *F. gummosa* essential oil (23).

In another study, Mahmoudvand and colleagues examined the apoptotic effects of three Iranian herbs on T. vaginalis. They found that Quercus infectoria (IC50=3.4 μ g/ml) was significantly more effective against the parasite than Satureja khuzestanica (IC50=5.1 μ g/ml) and Pistacia khinjuk (IC50=26.6 μ g/ml) (24).

Across studies reviewing the antitrichomonal potential of medicinal plants, the most active species typically belong to three families: Asteraceae, Lamiaceae, and Myrtaceae. Furthermore, terpenes, βglycosides, saponins, essential oils, and alkaloids are among the major phytochemical compounds responsible for antitrichomonal activity (25). Berberine, an isoquinoline alkaloid used traditionally as a natural antibiotic, is one of the main components of Argemone mexicana methanolic extract. The IC50 antitrichomonal activity of A. mexicana extract was 70.8 and 67.2 µg/ml for the stem and leaf, respectively (26). Other plants with strong antitrichomonal effects include Persea americana, Verbascum thapsus, and Ocimum basilicum (27). The IC50 values of P. americana seed extracts against T. vaginalis were 0.524 and 0.533 µg/ml for the chloroform and ethanolic extracts, respectively (28). Alcoholic extract of V. thapsus and O. basilicum essential oil were effective against T. vaginalis at 30 and 39.17 µg/ml, respectively, after 24 hours of exposure (29,30).

In the present study, the observed antitrichomonal properties of *F. pseudalliacea* may be attributed to the presence of bioactive components. Preliminary analysis demonstrated that coumarins, terpenoids, and steroids were the major bioactive constituents in the extracts. Phytochemical screening analyses are helpful for identifying bioactive compounds and supporting the discovery and development of medicinal agents. These studies also enable quantitative and qualitative evaluation of active pharmaceutical compounds in crude extracts.

One of the advantages of this study was the use of several *Trichomonas* isolates, which increases the reliability of the results. Additionally, essential oil and extracts were examined simultaneously, allowing a more accurate comparison of their effects. However, one of the main limitations of this study was the lack of access to metronidazole-resistant *Trichomonas* isolates, preventing evaluation of the compounds' effectiveness on resistant strains.

Conclusion

Despite limitations related to access to metronidazole-resistant isolates, *F. pseudalliacea* demonstrated potential anti-*Trichomonas* activity. The findings support the possibility of using components of *F. pseudalliacea* in the treatment of trichomoniasis. Therefore, further investigation into the bioactive components of *F. pseudalliacea* is recommended.

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Ethical statement

This study was approved by the Ethical Committee of Hamadan University of Medical Sciences, Hamadan, Iran (UMSHA.REC.1394.84 IR).

Conflicts of interest

The authors declare that they have no conflict of interests to disclose.

Author contributions

DD, MF, AHM, and MM designed the study. ZA, DD, and MM conducted the experiment and data collection. MM and DD analyzed and interpreted the data and prepared the manuscript. All authors read and approved the final manuscript for publication.

Data availability statement

The data that support the findings of this study are available from the corresponding author upon reasonable request.

References

- Schwebke JR, Burgess D. Trichomoniasis. Clin Microbiol Rev. 2004;17(4):794-803. [View at Publisher] [DOI] [PMID] [Google Scholar]
- Rowley J, Vander Hoorn S, Korenromp E, Low N, Unemo M, Abu-Raddad LJ, et al. Chlamydia, gonorrhoea, trichomoniasis and syphilis: global prevalence and incidence estimates, 2016. Bull World Health Organ. 2019;97(8):548-62. [View at Publisher] [DOI] [PMID] [Google Scholar]
- Robinson SC. Trichomonal vaginitis resistant to metronidazole. Can Med Assoc J. 1962;86(14):665. [View at Publisher] [PMID] [Google Scholar]
- Workowski KA, Berman SM. Centers for Disease Control and Prevention Sexually Transmitted Disease Treatment Guidelines. Clin Infect Dis. 2011;53(Suppl3):S59-63. [View at Publisher] [DOI] [PMID] [Google Scholar]
- 5. Nazari ZE, Iranshahi M. Biologically active sesquiterpene coumarins from Ferula species. Phytother Res. 2011;25(3):315-23. [View at Publisher] [DOI] [PMID] [Google Scholar]
- Abd El-Razek MH. Terpenoid coumarins of the genus Ferula. Heterocycles. 2003;34(22):689-716. [View at Publisher] [DOI] [Google Scholar]
- Bafghi AF, Bagheri SM, Hejazian SH. Antileishmanial activity of Ferula assa-foetida oleo gum resin against Leishmania major: An in vitro study. J Ayurveda Integr Med. 2014;5(4):223-6. [View at Publisher] [DOI] [PMID] [Google Scholar]
- 8. Khanmohammadi M, Ganji S, Rehyhani Rad S. Anti-protozoan Effects of Methanol Extracts of the Ferula szowitsiana on the Trichomonas Vaginalis Trophozoites in vitro. Int J Women's Health Reprod Sci. 2014;2(5):301-6. [View at Publisher] [DOI] [Google Scholar]
- 9. Dastan D, Salehi P, Gohari AR, Zimmermann S, Kaiser M, Hamburger M, et al. Disesquiterpene and sesquiterpene coumarins from Ferula pseudalliacea, and determination of their absolute configurations. Phytochemistry. 2012;78:170-8. [View at Publisher] [DOI] [PMID] [Google Scholar]
- Dastan D, Salehi P, Gohari AR, Ebrahimi SN, Aliahmadi A, Hamburger M. Bioactive sesquiterpene coumarins from Ferula pseudalliacea. Planta Med. 2014;80(13):1118-23. [View at Publisher] [DOI] [PMID] [Google Scholar]
- 11. Dastan D, Salehi P, Ghanati F, Gohari AR, Maroofi H, Alnajar N. Phytotoxicity and cytotoxicity of disesquiterpene and sesquiterpene coumarins from Ferula pseudalliacea. Ind Crops Prod. 2014;55:43-8. [View at Publisher] [DOI] [Google Scholar]
- 12. Dastan D, Salehi P, Aliahmadi A, Gohari AR, Maroofi H, Ardalan A. New coumarin derivatives from Ferula pseudalliacea with antibacterial activity. Nat Prod Res. 2016;30(24):2747-53. [View at Publisher] [DOI] [PMID] [Google Scholar]
- 13. Ugochukwu SC, Uche I A, Ifeanyi O. Preliminary phytochemical screening of different solvent extracts of stem bark and roots of Dennetia tripetala G. Baker. Asian J Plant Sci Res. 2013;3(3):10-3. [View at Publisher] [Google Scholar]
- Fathollahi R, Dastan D, Lari J, Masoudi S. Chemical composition, antimicrobial and antioxidant activities of Crupina crupinastrum as a medicinal plant growing wild in west of Iran. JRPS. 2018;7(2):174-82. [View at Publisher] [DOI] [Google Scholar]

- 15. Matini M, Rezaie S, Mohebali M, Maghsood A, Rabiee S, Fallah M, et al. Prevalence of Trichomonas vaginalis infection in Hamadan city, Western Iran. Iran J Parasitol. 2012;7(2):67-72. [View at Publisher] [PMID] [Google Scholar]
- Matini M, Maghsood AH, Mohebali M, Rabiee S, Fallah M, Rezaie S, et al. In vitro susceptibility of Iranian isolates of Trichomonas vaginalis to metronidazole. Iran J Parasitol. 2016;11(1):46-51.
 [View at Publisher] [PMID] [Google Scholar]
- 17. Schwebke JR, Barrientes FJ. Prevalence of Trichomonas vaginalis isolates with resistance to metronidazole and tinidazole. Antimicrob Agents Chemother. 2006;50(12):4209-10. [View at Publisher] [DOI] [PMID] [Google Scholar]
- 18. Ziaei Hezarjaribi H, Nadeali N, Fakhar M, Soosaraei M. Medicinal Plants with Anti-Trichomonas vaginalis Activity in Iran: A Systematic Review. Iran J Parasitol. 2019;14(1):1-9. [View at Publisher] [DOI] [PMID] [Google Scholar]
- Karami F, Dastan D, Fallah M, Matini M. In Vitro Activity of Foeniculum vulgare and Its Main Essential Oil Component Trans-Anethole on Trichomonas vaginalis. Iran J Parasitol. 2019;14(4):631-8. [View at Publisher] [DOI] [PMID] [Google Scholar]
- Akbari Z , Dastan D, Maghsood A H , Fallah M, Matini M. Investigation of In vitro Efficacy of Marrubium vulgare L. Essential Oil and Extracts Against Trichomonas vaginalis. Zahedan J Res Med Sci. 2018;20(9):e67003. [View at Publisher] [DOI] [Google Scholar]
- Matini M, Bakhtiyar nejad S, Dastan D, Maghsood A, Fallah M. Investigation of in-vitro efficacy of Pistacia atlantica subsp. kurdica extracts against Trichomonas vaginalis. Stud Med Sci. 2018;29(3):198-207. [View at Publisher] [Google Scholar]
- Matini M, Bakhtiarnejad S, Dastan D, Maghsood A H, Fallah M. In-Vitro Efficacy of Plantago lanceolata L. Extracts on Trichomonas Vaginalis. J Arak Uni Med Sci. 2017;20(6):74-82. [View at Publisher] [Google Scholar]
- Akbari M, Dastan D, Fallah M, Matini M. In-vitro Activity of Ferula gummosa Essential Oil and Its Different Extracts on Trichomonas vaginalis. Sjimu. 2019;27(2):1-10. [View at Publisher] [DOI] [Google Scholar]
- 24. Mahmoudvand H, Badparva E, Baharvand Z, Salehi Lalehmarzi H. Anti-Trichomonas vaginalis activities and apoptotic effects of some Iranian medicinal plants. Trop Biomed. 2018;35(2):347-53. [View at Publisher] [PMID] [Google Scholar]
- Hashemi N, Ommi D, Kheyri P, Khamesipour F, Setzer WN, Benchimol M. A review study on the anti-trichomonas activities of medicinal plants. Int J Parasitol Drugs Drug Resist. 2021;15:92-104. [View at Publisher] [DOI] [PMID] [Google Scholar]
- Elizondo-Luevano JH, Verde-Star J, González-Horta A, Castro-Ríos R, Hernández-García ME, Chávez-Montes A. In Vitro Effect of Methanolic Extract of Argemone mexicana against Trichomonas vaginalis. Korean J Parasitol. 2020;58(2):135-45. [View at Publisher] [DOI] [PMID] [Google Scholar]
- 27. Mehriardestani M, Aliahmadi A, Toliat T, Rahimi R. Medicinal plants and their isolated compounds showing anti-Trichomonas vaginalis-activity. Biomed Pharmacother. 2017;88:885-93. [View at Publisher] [DOI] [PMID] [Google Scholar]
- Jiménez-Arellanes A, Luna-Herrera J, Ruiz-Nicolás R, Cornejo-Garrido J, Tapia A, YépezMulia L. Antiprotozoal and antimycobacterial activities of Persea americana seeds. BMC Complement Altern Med. 2013;13:109. [View at Publisher] [DOI] [PMID] [Google Scholar]
- 29. Ezz Eldin HM, Badawy AF. In vitro anti-Trichomonas vaginalis activity of Pistacia lentiscus mastic and Ocimum basilicum essential oil. J Parasit Dis. 2015;39(3):465-73. [View at Publisher] [DOI] [PMID] [Google Scholar]
- Kashan ZF, Arbabi M, Delavari M, Hooshyar H, Taghizadeh M, Joneydy Z. Effect of Verbascum thapsus ethanol extract on induction of apoptosis in Trichomonas vaginalis in vitro. Infect Disord Drug Targets. 2015;15(2):125-30. [View at Publisher] [DOI] [PMID] [Google Scholar]

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