



## Investigating the role of miR-30c in modulating $\gamma$ -secretase activity: Implications for Alzheimer's disease

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### Abstract

**Background:** Alzheimer's disease (AD), the most common form of dementia, affects millions of people worldwide. Clinical trials using anti-A $\beta$  antibodies demonstrate that amyloid plaque removal in early-stage AD can slow disease progression. Along with  $\beta$ -secretase,  $\gamma$ -secretase plays a role in cleaving amyloid precursor protein (APP). The aim of this study was to use computational docking to identify molecules that can activate  $\gamma$ -secretase.

**Methods:** Initially, the targets of hsa-miR-30c-5p were assessed using the TargetScanHuman server. The structure of  $\gamma$ -secretase was prepared in Chimera by removing non-standard residues and water molecules. Adjacent amino acids to the cholesterol ligand were then identified using PyMOL. The 3D structure and SMILES notation for cholesterol were obtained from PubChem. Docking results in pdbqt format were analyzed using Discovery Studio, LigPlus+, and PDBsum, with LigPlus+ focusing on protein subunit interactions.

**Results:** The TargetScanHuman server indicated that  $\gamma$ -secretase is a target of hsa-miR-30c-5p. Drug-like properties (Solubility, tumorigenicity, LogP, toxicity) of compounds were predicted using tools such as SwissTargetPrediction, PASS-Way2Drug, and SwissADME, following Lipinski's Rule of Five. Amino acids Trp227, Leu192, Arg186, Leu199, Leu203, Leu206, Tyr155, Leu215, Phe162, Ser223, and Ile230, located on the  $\gamma$ -secretase C subunit, were analyzed for interactions using LigPlot after AutoDock Vina docking and Chimera visualization.

**Conclusion:** These in silico findings suggest cholesterol acetate as a potential activator of  $\gamma$ -secretase; further experimental validation is warranted.

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### Introduction

Alzheimer's disease is the most prevalent form of dementia globally, affecting millions of individuals. Until recently, treatments for Alzheimer's disease have primarily focused on managing symptoms rather than addressing underlying causes. Memantine, approved about 20 years ago, is a notable example of such symptomatic treatments aimed at alleviating disease manifestations. The amyloid cascade hypothesis proposes that the accumulation of amyloid-beta (A $\beta$ ) in the brain is the primary trigger for Alzheimer's disease (1).

Among the dysregulated microRNAs in AD, miR-30c directly targets PSEN2, a catalytic component of  $\gamma$ -secretase, potentially reducing enzyme activity and altering A $\beta$  processing. Therefore, it has been hypothesized that small-molecule activation of  $\gamma$ -secretase could compensate for miR-30c-mediated suppression (2,3). A persistent imbalance between the production and clearance of amyloid-beta (A $\beta$ ) can result in elevated levels of A $\beta$ 42 (4).

Analysis of microRNAs obtained from small RNA sequencing of blood samples from elderly patients with Alzheimer's disease (Average age: 70.3  $\pm$  7.9 years) using the tool "omiRas" showed an elevation in miR-30c-5p. Similarly, in a rabbit model of late-onset Alzheimer's disease (LOAD) fed a cholesterol-enriched diet, increased levels of miR-30c were observed in the cerebral cortex (2).

The  $\gamma$ -secretase complex is a transmembrane protein assembly composed of four key components: Presenilin (PS), nicastrin, anterior pharynx defective-1 (Aph-1), and presenilin enhancer-2 (Pen-2).  $\gamma$ -Secretase is classified as an intramembrane-cleaving protease (I-CLiP), a distinct category of enzymes that cleave substrates within the lipid bilayer of cell membranes (5,6).  $\gamma$ -Secretase, together with  $\beta$ -secretase, processes amyloid precursor protein (APP) through sequential

cleavages. After  $\beta$ -secretase generates the C99 fragment,  $\gamma$ -secretase performs the final intramembrane cleavage, releasing A $\beta$  peptides (e.g., A $\beta$ 40, A $\beta$ 42) and the APP intracellular domain (AICD) (7-10).

In vitro studies involving human umbilical vascular endothelial cells (HUVECs) have demonstrated that miR-30c-5p can reduce inflammatory responses, including activation of nuclear factor kappa light-chain-enhancer of activated B cells (NF- $\kappa$ B) and oxidative stress induced by oxidized low-density lipoprotein (3).

The aim of this study was to use computational methods, specifically molecular docking, to identify molecules capable of activating the  $\gamma$ -secretase enzyme. Using Molegro Virtual Docker, group docking was performed on 10 cholesterol-derived molecules, and the results were analyzed. SwissADME was subsequently used to evaluate the chemical properties and toxicity of the identified molecules.

### Methods

#### Identification of targets of hsa-miR-30c-5p

Initially, all targets of miR-30c were identified using the TargetScanHuman server ([https://www.targetscan.org/vert\\_80/](https://www.targetscan.org/vert_80/)). Targets involved in Alzheimer's disease were identified using the KEGG server (<https://www.kegg.jp/kegg/pathway.html>). TargetScanHuman v8.0 (September 2021) showed a cumulative weighted context++ score of -0.73.

#### Preparation of proteins and ligands

The three-dimensional structure of the  $\gamma$ -secretase enzyme was obtained from the Protein Data Bank (PDB entry code: 8k8e) with suitable resolution. The TargetScanHuman server identified the presenilin 2 gene as a predicted target of miR-30c. The  $\gamma$ -secretase structure was prepared

using Chimera software by removing non-standard structures and water molecules, followed by energy minimization. Cholesterol (PubChem ID: 5997), a known stimulator of  $\gamma$ -secretase, was isolated from the protein structure and energy-minimized using ChemBio3D software. The amino acids adjacent to the cholesterol ligand were mapped using PyMOL software. Subsequently, the energy-minimized structures of both the enzyme and ligand were imported into Chimera and VMD for further analysis. The role of the ligand was examined using the Way2Drug and SwissTargetPrediction web servers, which revealed that cholesterol functions as a stimulator of the enzyme.

### Molecular docking

The 3D structure and SMILES notation of cholesterol (PubChem CID: 5997) were retrieved from the PubChem database. A dataset of 100 cholesterol-containing molecules was obtained from the ZINC15 server, from which 10 compounds were selected for molecular docking analysis. These molecules underwent group docking in Molegro Virtual Docker to determine their optimal binding conformations. Chimera was used to pre-optimize the protein structure, and energy-scored group docking narrowed the 10 ligands to one optimal activator. Final docking was performed in Chimera using AutoDock Vina (Version 1.5.6) with the selected ligand, which had been optimized in ChemBio3D software (Total energy: 24.4520 kcal/mol). Preparation of the protein structure involved removing water molecules and non-standard residues, followed by the addition of polar hydrogen atoms. Subsequent calculations of atomic charges, solvation parameters, and component volumes were performed using AutoDock (Steepest descent steps: 200; steepest descent step size (Å): 0.02; conjugate gradient steps: 10; conjugate gradient step size (Å): 0.02; update interval: 100). The final file was formatted in pdbqt format, containing partial atomic charges and atom types (Center: -34.028, -64.0541, 45.1775; size: 27.2784, 15.8822, 24.7592). Docking results were analyzed using Discovery Studio and LigPlus+, as well as online servers such as PDBsum and the ligand-protein interaction profile server.

### Prediction of physicochemical and biological properties of studied compounds

Since suitable physicochemical properties are crucial for a ligand, this study evaluated key parameters including water solubility, tumorigenicity, LogP value, and toxicity using databases such as SwissTargetPrediction, PASS-Way2Drug, and SwissADME, in accordance with Lipinski's Rule of Five. Lipinski's Rule of Five is a concept frequently used in drug discovery and is based on pharmacokinetic properties such as absorption, distribution,

metabolism, and excretion, which depend on specific physicochemical criteria: No more than 5 hydrogen bond donors, no more than 10 hydrogen bond acceptors, molecular mass less than 500 Da, and a partition coefficient not greater than 5. LogP is an important component of Lipinski's Rule of Five and predicts the drug-likeness of a new compound. According to this rule, an oral drug should have a LogP value < 5, ideally between 1.35 and 1.8 for good oral and intestinal absorption. There is no fixed threshold value for toxicity in SwissADME; in general, ligand toxicity is inferred based on inhibition of CYP1A2, CYP2C19, CYP2C9, CYP2D6, and CYP3A4. In PASS online analysis, a PA value > 0.3 is typically considered indicative of ligand-mediated inhibition or activation of a target.

## Results

### Identification of targets of hsa-miR-30c-5p

All targets of miR-30c were identified using the TargetScanHuman server, and targets involved in Alzheimer's disease were recognized through the KEGG server (Table 1). It is important to note that PSEN2 is a subunit of the  $\gamma$ -secretase enzyme.

### Active-site validation (Cholesterol re-docking)

Initially, the cholesterol ligand was removed from the enzyme's active site. Its role as a stimulator of the  $\gamma$ -secretase enzyme was confirmed using SwissTargetPrediction servers, and subsequent docking results were compared. PyMOL software was used to identify amino acids surrounding the active site, specifically those within a 4-angstrom radius (Figure 1). The identified amino acids - Trp227, Leu192, Arg186, Leu199, Leu203, Leu206, Tyr155, Leu215, Phe162, Ser223, and Ile230 - were located on the C subunit of the  $\gamma$ -secretase enzyme.

### Virtual screening outcome (Ranking of 10 sterols)

The cholesterol structure was imported into the ZINC15 database, allowing the extraction of a dataset comprising 103 molecules. From this dataset, 10 molecules were selected for further analysis using group docking in Molegro Virtual Docker. The molecule with the most favorable energy values was selected as the best candidate for binding to the active site and was identified as cholesterol acetate (Table 2). The more negative the MolDock and reRank scores, the stronger the ligand binding. Cholesterol acetate exhibited suitable scores for both metrics, indicating effective interaction with the macromolecule. Using PyMOL software, amino acids within a 4-angstrom radius of the ligand were identified (Figure 2). These amino acids included Phe682, Val686, Thr687, Leu206, Leu196, Phe698, Phe229, Phe173, Gly234, Gln116, Ala232, Val176, Arg115, and Tyr119.

Table 1. Targets of hsa-miR-30c

Gene	Representative transcript	Gene name	Number of 3P-seq tags supporting UTR + 5
ADAM10	0260408.3	ADAM metalloproteinase domain 10	385
CABLES2	0279101.5	Cdk5 and Abl enzyme substrate 2	450
PSEN2	0340188.4	Presenilin 2 (Alzheimer disease 4)	613
CAPN5	0531028.1	Calpain 5	89

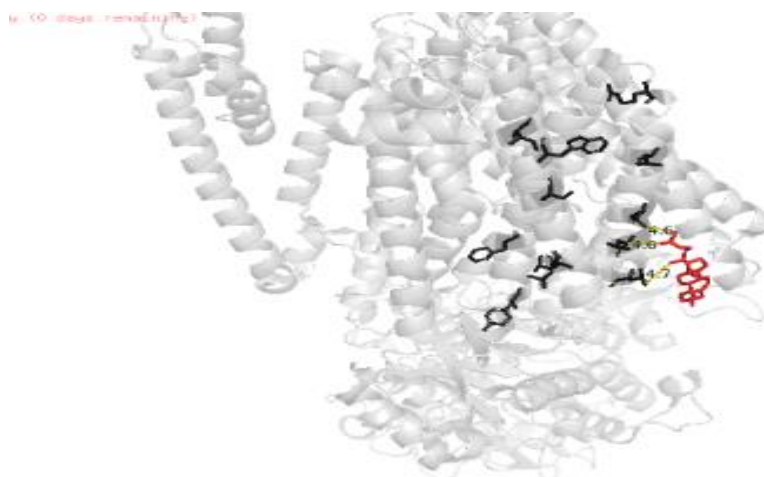
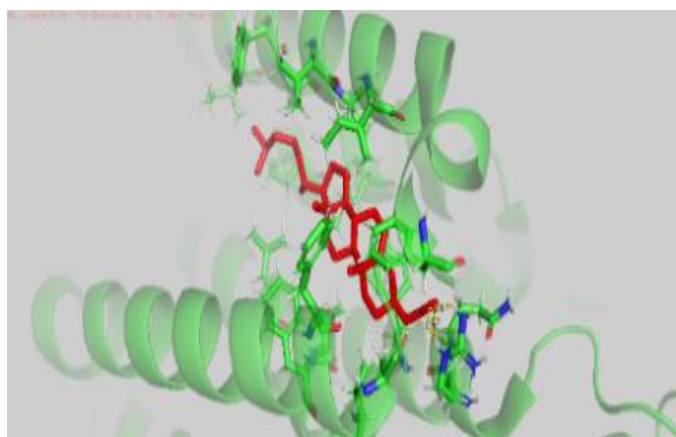


Figure 1. Amino acids close to cholesterol in the C chain

**Table 2.** Binding energy and interactions between the studied compounds and the amino acids of the active site of the  $\gamma$ -secretase enzyme

Molecule number	Molecule name	MolDock score	ReRanke score	Hydrogen bond	Hydrophobic bond
1	Cerebrosterin	-64/03	-29/56	Val103 (B), Arg652 (A), Thr188 (B)	Val103 (B), Arg652 (A), Thr188 (B), Phe105 (B), Lys187 (B), Glu184 (B), Glu245 (A)
2	22b-Hydroxycholesterol	-83/00	-36/76	Val103 (B), Glu184 (B), Thr188 (B), Ile246 (A)	Lys654 (A), Glu184 (B), Lys187 (B)
3	(25s)-26-Hydroxycholesterol	-73/63	-18/45	Pro244 (A), Lys187 (B)	Pro244 (A), Lys187 (B), AB), Thr188 (B), Arg108 (B), Asn243 (A), Glu184 (B), Asn109 (B)
4	Avenasterol	-76/07	-37/85	Ser56 (A)	Lys654 (A), His220 (A), Gly68 (A), Asp655 (A), Ser67 (A), Thr107 (B), Arg108 (B), Lys654 (A)
5	24 (r)-Hydoxycholesterol	-69/97	-30/81	Glu184 (B), Ser67 (A), Gly68 (A)	Ile66 (A), Asp655 (A)
6	Cholesterin Acetate	-85/52	-43/83	0	Lys187 (B), Arg108 (B), Ile66 (B), Thr188 (B), Phe105 (B), Glu184 (B)
7	Cholesterol methyl ether	-80/75	11/50	Thr188 (B)	Ile66 (A), His220 (A), Asp655 (A), Phe218 (A), Lys654 (A), Thr188 (B), Glu184 (B)
8	Cholesterin ethyl ether	-82/99	-32/38	Arg108 (B)	Lys187 (B), Thr188 (B), Arg108 (B), Asn243 (A), Phe105 (B), Lys654 (A)
9	7-Hydroxycholesterol	-82/99	-32/38	Ile246 (A), Arg652 (A)	Arg652 (A), Ile246 (A), Glu184 (B), Thr188 (B), Val103 (B)
10	Campesterol	-82/99	-32/38	0	Ile66 (A), Phe218 (A), His220 (A), Glu184 (B), Lys654 (A), Asp655 (A), Ser219 (A)
11	Cholesterol	-79/92	-31/00	Asp655 (A)	Phe218 (A), Ile66 (A), Thr188 (B), Lys654 (A)

**Figure 2.** Amino acids around the cholesterol acetate ligand**ADMET profile (SwissADME/PASS results)**

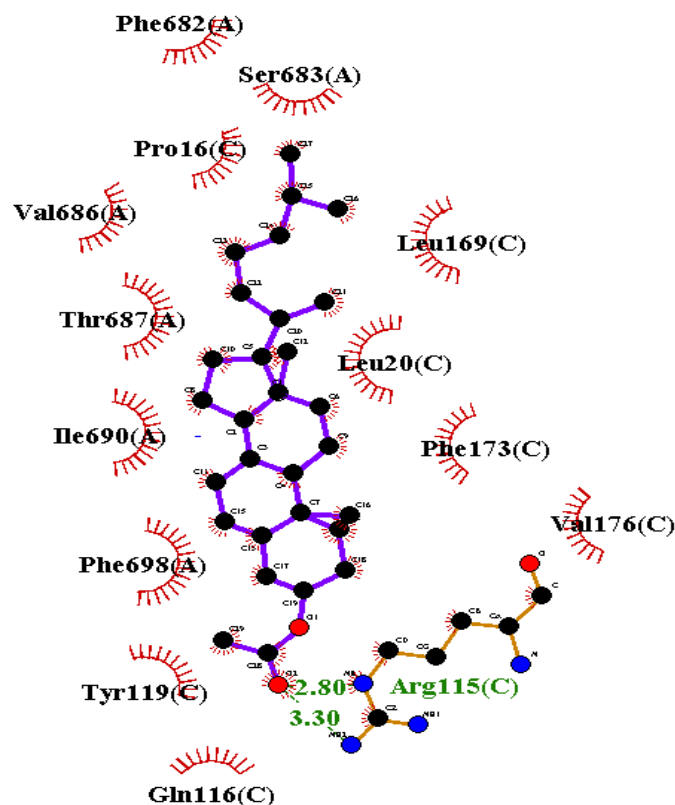
In accordance with Lipinski's Rule of Five, this study evaluated key physicochemical properties - solubility, tumorigenicity, LogP, and toxicity - of the compounds using predictive tools such as SwissTargetPrediction, PASS-Way2drug, and SwissADME. According to Lipinski's Rule of Five, the LogP value, representing the logarithm of the octanol/water partition coefficient, serves as an indicator of a compound's solubility in both water and fat and effectively acts as a solubility index. A ligand with low hydrophilicity exhibits reduced absorption. According to the SwissADME database, values above 15.4 are considered acceptable under Lipinski's Rule of Five, and all studied ligands met this criterion, as shown in [Table 3](#).

**Detailed binding analysis (LigPlot; Interaction geometry)**

After docking of the selected ligand, cholesterolin acetate, using AutoDock Vina within the Chimera platform, the optimal conformation was obtained and saved in PDB format. The docking results were analyzed and evaluated using LigPlot software ([Figure 3](#)).

**Table 3.** Results from the toxicity risk assessment of designed cholesterol-based ligands

Molecular number	Toxicity	LogP	Solubility	Molecular weight (g/mol)	Crossing the blood-brain barrier	Hydrogen bond donor	Hydrogen bond acceptor
1	-	5/41	Weak	402/65	-	2	2
2	-	5/41	Weak	402/65	-	2	2
3	-	5/41	Weak	402/65	-	2	2
4	-	6/62	Weak	412/69	-	1	1
5	-	5/41	Weak	402/65	-	2	2
6	-	6/51	Weak	428/69	-	0	2
7	-	6/54	Weak	400/68	-	0	1
8	-	6/73	Weak	414/71	-	0	1
9	-	5/41	Weak	402/65	-	2	2
10	-	6/54	Weak	400/68	-	1	1
11	-	6/34	Weak	386/65	-	1	1



**Figure 3.** Investigation of the amino acids involved in the formation of hydrogen and hydrophobic bonds between cholesterol acetate and the  $\gamma$ -secretase enzyme (Green dashed lines indicate hydrogen bonds, and brackets indicate hydrophobic bonds)

## Discussion

Previous research has explored the expression patterns of  $\gamma$ -secretase complex subunits in the brain and spinal cord, indicating that  $\gamma$ -secretase plays a critical role in activating apoptosis within sympathetic neurons during the postnatal stage in rats (7,9,10). Sensitive biochemical and imaging biomarker technologies have made it feasible to observe A $\beta$  amyloidosis as the disease advances. Research indicates that the onset of A $\beta$  amyloidosis occurs approximately 10 - 15 years prior to symptom presentation in both sporadic and familial Alzheimer's disease (11,12).  $\gamma$ -Secretase modulators (GSMs) are compounds that influence the activity of the  $\gamma$ -secretase complex, which is crucial in the processing of amyloid precursor protein (APP) into amyloid-beta (A $\beta$ ) peptides. One of the primary mechanisms by which GSMs operate is by shifting the  $\epsilon$ -cleavage of APP to favor the production of shorter A $\beta$  species, particularly A $\beta$ 38 and A $\beta$ 40, rather than the longer and more aggregation-prone A $\beta$ 42. This modulation is significant because the accumulation of A $\beta$ 42 is closely associated with the development of Alzheimer's disease (13,14).

In examining the docked pose of cholesterol acetate, it is essential to determine whether it occupies the same allosteric pocket as other known GSMs. If cholesterol acetate does indeed bind to this allosteric site, it may enhance the preferential cleavage of APP toward shorter A $\beta$  species (Table 2). This interaction could theoretically impact the A $\beta$ 42/A $\beta$ 40 ratio by decreasing the production of A $\beta$ 42 while increasing the relative abundance of A $\beta$ 40, thereby potentially reducing the overall pathogenicity associated with A $\beta$ 42 accumulation (15). Moreover, the overall catalytic turnover of the  $\gamma$ -secretase complex could be affected by this modulation. Structural studies and kinetic analyses have shown that alterations in the binding dynamics of GSMs can lead to changes in the efficiency of substrate processing. For instance, the literature indicates that specific allosteric interactions can enhance or inhibit the catalytic activity of  $\gamma$ -secretase, which in turn influences the balance of A $\beta$  species produced. This relationship highlights the importance of understanding the binding characteristics of cholesterol acetate within the context of  $\gamma$ -secretase modulation.

Because miR-30c lowers PSEN2 translation by approximately 40% in neuronal models, a ligand that increases  $\gamma$ -secretase activity by at least 50% would, in theory, restore net enzymatic flux. Molecular dynamics analyses indicate that cholesterol acetate stabilizes the open conformation of PSEN2 catalytic aspartates, potentially increasing  $k_{cat}$ . In 2001, a groundbreaking study, introduced the concept of GSMs as a novel approach to regulating the production of amyloid-beta (A $\beta$ ) peptides through  $\gamma$ -secretase. This discovery provided an alternative strategy for influencing A $\beta$  production, which is crucial in the context of Alzheimer's disease (16,17). Studies further indicate that the  $\gamma$ -secretase complex contributes to neurite outgrowth in central nervous system neurons, particularly affecting axonal growth and dendritic spine development (7,9,11,12). The  $\gamma$ -secretase complex subunits demonstrate widespread expression across tissues, observable at both transcriptional (mRNA) and translational (Protein) levels. To elucidate their physiological roles, researchers have employed knockout (KO) mouse models to systematically analyze subunit functionality (7,18,19). In Alzheimer's disease research, the role of statins as a treatment approach continues to be debated, with no conclusive consensus established to date (20,21).

An analysis of 22,000 medical records indicated that patients using lovastatin or pravastatin for cardiac conditions showed significantly reduced rates of Alzheimer's disease diagnosis compared with counterparts receiving non-statin cardiac therapies. Consequently, despite robust evidence implicating cholesterol - especially ApoE4-associated pathways - as a risk factor for Alzheimer's disease, the therapeutic potential of cholesterol modulation in AD treatment remains ambiguous (22,23). In summary, despite being an endogenous metabolite, cholesterol acetate acts as a potent modulator of  $\gamma$ -secretase (24-26). Research has also revealed that nonsteroidal anti-inflammatory drugs (NSAIDs) such as ibuprofen, indomethacin, and sulindac sulfide regulate  $\gamma$ -secretase, acting as pioneering carboxylic acid-derived  $\gamma$ -secretase modulators (27).

## Conclusion

Treating and preventing Alzheimer's disease remains a challenging task. However, the use of anti-amyloid-beta (A $\beta$ ) antibodies has revitalized the field by demonstrating substantial clinical improvements. Our in-silico data identify cholesterol acetate as a candidate  $\gamma$ -secretase activator; however, empirical validation is required before its therapeutic relevance can be assessed. Cholesterol acetate showed the most favorable Vina binding energy ( $-10.3 \text{ kcal mol}^{-1}$ ) and no predicted PAINS or toxicity alerts, and enzyme-based confirmation assays are currently being initiated.

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## Ethical statement

The present study was approved under the ethical approval number IR.IAU.TABRIZ.REC.1401.094 at the Islamic Azad University, Tabriz, Iran.

## Conflicts of interest

The authors declare that they have no competing interests to report.

## Author contributions

T. K. conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the article, and approved the final draft. R. S. performed the experiments, authored or reviewed drafts of the article, and approved the final draft. S. F. reviewed drafts of the article, supervised, and approved the final draft. S. N. conceived and designed the experiments, analyzed the data, authored or reviewed drafts of the article, supervised, and approved the final draft.



## Data availability statement

The data that support the findings of this study are available from the corresponding author upon reasonable request.

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